

P-ISSN: 2349–8528 E-ISSN: 2321–4902 www.chemijournal.com IJCS 2020; 8(3): 1317-1321 © 2020 IJCS Received: 10-03-2020 Accepted: 12-04-2020

V Sravani

Department of Vegetable Science, Navsari Agricultural University, Navsari, Gujarat, India

SN Saravaiya

Department of Vegetable Science, Navsari Agricultural University, Navsari, Gujarat, India

DR Bhanderi

Department of Vegetable Science, Navsari Agricultural University, Navsari, Gujarat, India

Dev Raj

Department of Vegetable Science, Navsari Agricultural University, Navsari, Gujarat, India

Himani B Patel

Department of Vegetable Science, Navsari Agricultural University, Navsari, Gujarat, India

JM Vashi

Department of Vegetable Science, Navsari Agricultural University, Navsari, Gujarat, India

Corresponding Author: V Sravani Department of Vegetable Science, Navsari Agricultural University, Navsari, Gujarat, India

Effect of plant growth regulators on biochemical analysis of onion (Allium cepa)

V Sravani, SN Saravaiya, DR Bhanderi, Dev Raj, Himani B Patel and JM Vashi

DOI: https://doi.org/10.22271/chemi.2020.v8.i3r.9381

Abstract

A field experiment was carried out during Rabi 2018 and 2019 at RHRS, Navsari Agricultural University, Navsari, Gujarat to study the effect of different plant growth regulators like GA₃ and NAA on biochemical parameters of onion. The treatment T_1 (GA₃ 25 mg l⁻¹) recorded highest chlorophyll content of leaves 45 DATP (1.06 mg 100 g⁻¹), 60 DATP (2.64 mg 100 g⁻¹), 90 DATP (2.32 mg 100 g⁻¹), ascorbic acid (10.45 mg 100 g⁻¹), phenols (61.77 mg 100 g⁻¹) and proteins (1072.54 mg 100 g⁻¹) and found significant. The moisture content (83.98%) under T₆ (NAA 75 mg l⁻¹) were found significant.

Keywords: Plant growth regulators, GA₃, NAA, onion etc.

Introduction

India is the world's second largest producer of vegetables (187.47 million tonnes) next only to China (Anonymous, 2019). Onion (*Allium cepa* L.) is an important and indispensable item in every kitchen as condiment cum vegetable in India. It is one of the important underground bulbous vegetable crops of Alliaceae family and is said to be native of Central Asia and Mediterranean region (Mc Collum, 1976). Plant growth regulators are organic compounds other than nutrients which in small amount promotes / inhibit or otherwise modify any physiological response in plant (Purohit, 2007)^[9]. Plant bioregulators called as magic chemicals are new generation agrochemicals, when added in small quantity, modify the natural growth regulatory systems right from seed germination to senescence in several vegetable crops and also regulate and modify various physiological processes within the plant and they help to increase the yield (Weaver, 1972)^[12].

Materials and methods

The field experiment was carried out at the vegetable research scheme, Regional Horticultural Research Station of the Navsari Agricultural University, Navsari, Gujarat, India during *Rabi* 2018 and 2019 on cv. Gujarat Junagadh Red Onion 11 to investigate the response of plant bioregulators on growth parameters and plant growth analysis of onion. The experiment was conducted in Randomized Block Design (RBD) with three replications, which included 12 treatments namely, T₁: GA₃ 25 mg l⁻¹, T₂: GA₃ 50 mg l⁻¹, T₃: GA₃ 75 mg l⁻¹, T₄: NAA 25 mg l⁻¹, T₅: NAA 50 mg l⁻¹, T₆: NAA 75 mg l⁻¹, T₇: GA₃ 25 mg l⁻¹ + NAA 25 mg l⁻¹, T₈: GA₃ 25 mg l⁻¹ + NAA 50 mg l⁻¹, T₉: GA₃ 25 mg l⁻¹ + NAA 75 mg l⁻¹, T₁₀: GA₃ 50 mg l⁻¹ + NAA 50 mg l⁻¹, T₁: GA₃ 75 mg l⁻¹ + NAA 50 mg l⁻¹, T₁₀: GA₃ 75 mg l⁻¹ + NAA 50 mg l⁻¹, T₁₀: GA₃ 75 mg l⁻¹ + NAA 50 mg l⁻¹, T₁₀: GA₃ 75 mg l⁻¹ + NAA 50 mg l⁻¹, T₁₀: GA₃ 75 mg l⁻¹ + NAA 50 mg l⁻¹, T₁₀: GA₃ 75 mg l⁻¹ + NAA 75 mg l⁻¹ + NAA 75 mg l⁻¹ + NAA 50 mg l⁻¹, T₁₀: GA₃ 75 mg l⁻¹ + NAA 75 mg l⁻¹ and T₁₂: Control. The foliar sprays were made at 30 days after transplanting during morning hours to avoid the dehydration effect. For recording different observations, ten plants of onion from each net plot area were selected randomly and tagged with labels.

Results

Proteins (mg 100 g⁻¹)

Results of proteins in onion bulb under different treatments showed significant in pooled analysis. The maximum protein content (1072.54 mg) observed with T_1 (GA₃ 25 mg l⁻¹) followed by the treatment T_{10} (GA₃ 50 mg l⁻¹ + NAA 50 mg l⁻¹). Whereas, GA₃ 75 mg l⁻¹ + NAA 75 mg l⁻¹ (T₁₁) recorded minimum protein content (866.82 mg). The interaction of year ×

treatment was found non-significant.

Phenols (mg 100 g⁻¹)

The results of percent total phenols under different treatments showed significant in pooled analysis. The maximum phenol content (61.77 mg) observed with the treatment T_1 (GA₃ 25 mg l⁻¹) which was significantly at par with T_{12} . The minimum phenol content (51.53 mg) observed with the treatment T_3 . The interaction of year × treatment was found non-significant.

Chlorophyll content of leaves (mg 100g⁻¹)

The results of chlorophyll content under different treatments found significant. In pooled analysis, the maximum chlorophyll content (1.06 mg; 2.64 mg and 2.32 mg) at 45, 60 and 90 DATP respectively was recorded with the treatment T_1 (GA₃ 25 mg l⁻¹). The minimum chlorophyll content (0.82 mg) was recorded with T_{12} (Control) at 45 DATP whereas, chlorophyll content (2.33 mg and 2.07 mg) at 60 and 90 DATP respectively, was observed with the treatment T_3 (GA₃ 75 mg l⁻¹).

Reducing sugar (%)

During the first season, the maximum percentage of reducing sugars (5.21%) observed with T₄ (NAA 25 mg l⁻¹) which was significantly at par with the treatments T₅, T₉, T₆ and T₃. Whereas, the minimum reducing sugars (3.67%) was registered with control (T₁₂). During the second season, the treatment GA₃ 75 mg l⁻¹ (T₃) was recorded maximum reducing sugars percentage (6.62%). Whereas, T₄ (NAA 25 mg l⁻¹) recorded minimum reducing sugars percentage (4.15%). In pooled analysis, the results of reducing sugars under different treatments found non-significant.

Total sugar (%)

During the first season, the maximum total sugar (6.94%) was found with the treatment T_{10} (GA₃ 50 mg l⁻¹+ NAA 50 mg l⁻¹) which was significantly at par with the treatment T_8 . As well as, the minimum percent of total sugars (5.06%) was found with the treatment control (T_{12}). During the second season, application of NAA 75 mg l⁻¹ (T_6) recorded maximum total sugar (8.65%) which was significantly at par with the treatment T_9 . Whereas, GA₃ 25 mg l⁻¹ (T_1) recorded minimum total sugars (5.73%). Looking to the mean of pooled analysis, results of total sugars under different treatments was nonsignificant.

Non- reducing sugar (%)

During the first season, maximum non-reducing sugars (2.82%) found with the treatment T_{10} (GA₃ 50 mg l⁻¹ + NAA 50 mg l⁻¹) which was superior over other treatments followed by T_8 . Whereas, the minimum non-reducing sugars (0.59%) was found with the treatment T_4 (NAA 25 mg l⁻¹). The significantly maximum non-reducing sugars (4.76%) was observed with the treatment T_6 (NAA 75 mg l⁻¹) followed by T_9 during the second season. Whereas, the minimum non-reducing sugars (0.73%) was recorded with the treatment T_3 (GA₃ 75 mg l⁻¹). The results of percent non-reducing sugars under different treatments showed non-significant in pooled analysis. The interaction of year × treatment was found non-significant.

TSS (%)

In pooled analysis, the results of TSS under different treatments showed non-significant. The interaction of year \times treatment was found significant.

Moisture content (%)

Looking to the mean of pooled analysis, the maximum moisture content (83.98%) was obtained with T_6 (NAA 75 mg l⁻¹) which was significantly at par with the treatments T_5 , T_8 , T_{11} , T1 and T_{10} . The minimum moisture content (78.81%) was noted in control (T_{12}). The interaction of year × treatment was found non-significant.

Ascorbic acid content (mg 100 g⁻¹)

The results of ascorbic acid under different treatments showed significant in pooled analysis. The maximum ascorbic acid content (10.45 mg) was observed with the treatment T_1 (GA₃ 25 mg l⁻¹) which was significantly superior followed by T_{10} . The minimum ascorbic acid content (8.16 mg) was observed with the treatment control (T_{12}). The interaction of year × treatment was found non-significant.

Bulb pH

The significantly highest bulb pH (4.78) was found with the application of GA₃ 75 mg l⁻¹ + NAA 75 mg l⁻¹ (T₁₁) which was significantly at par with the treatments T₃, T₁₂, T₆, T₉, T₈, T₅, T₁₀, T₂ and T₄ in first season. Whereas the lowest bulb pH (4.48) was obtained in T₁ (GA₃ 75 mg l⁻¹). During the second season, the maximum bulb pH (4.77) was observed with T₆ (NAA 75 mg l⁻¹) which was significantly at par with the other treatments T₅, T₃, T₁₁, T₁₀, T₇, T₁, T₂ and T₄. The minimum bulb pH (4.38) was observed in control (T₁₂). The results of bulb pH under different treatments varied from 4.55 to 4.76 but it was non-significant. The interaction of year × treatment was found significant

Discussion

Chlorophyll-a and Chlorophyll-b were increased with GA₃ treatments in Andrographis paniculata (Gomatinayagam et al., 2009). The results indicated that there was increase in chlorophyll content due to foliar application of gibberellic acid (GA₃) induces enhancement of ultra structural morphogenesis of plastids, which coupled with retention of chlorophyll, delay plant senescence (Arteca, 1996; Ouzounidu and Ilias, 2005; Shah et al., 2007; Ouzounidu et al., 2011)^{[1, 6,} ^{10]}. This increase undoubtedly might have helped to improve the photosynthetic efficiency. The application of growth regulators may prove beneficial for improvement of growth and productivity of economically important vegetable crop onion. The plants under the influence of GA₃ was found with increased soluble carbohydrates, ascorbic acid content in tomato. Growth regulators improve the quality parameters of the onion bulbs due to enhanced physiological activity. The increase in TSS may be accounted due to the hydrolysis of polysaccharides. Conversion of organic acids in to soluble sugars and enhanced solubilisation of insoluble starch and pectin present in the cell wall and middle lamella. The increase in TSS content due to growth regulators found from the results of several workers (Tiwari et al., 2003; and Patel et al., 2010) [11, 8] and Govind et al. 2015 [4] in garlic. The production of ascorbic acid content seems to be enhanced under GA₃. The increase in ascorbic acid content may be due to vitamin can be synthesized in plant by the process involves the conversion of hexose sugar mainly glucose and galactose into ascorbic acid. The results are in conformity with Ouzounidou et al. (2011)^[5] in onion, Chaudhary et al. (2006) ^[2] and Ouzounidu et al. (2010) ^[7] in chilli and pepper. The moisture content showed significant results with NAA. Similar results with Govind et al. (2015)^[4] in garlic.

Acknowledgements

I wish to express my sincere gratitude to Dr. S.N. Saravaiya, Associate Professor for providing an opportunity to do my

project work in RHRS, Navsari Agricultural University, Gujarat during the year 2018 - 2019.



Fig 1: Effect of different treatments on phenol content (mg 100 g⁻¹) in onion



Fig 2: Effect of different treatments on protein content (mg 100 g⁻¹) in onion

	Chlorophyll content of leaves (mg 100 g ⁻¹)									
Treatments		45 DA'	ГР		60 DATP			90 DATP		
	2018	2019	Pooled	2018	2019	Pooled	2018	2019	Pooled	
T ₁ : GA ₃ 25 mg l ⁻¹	1.08	1.04	1.06	2.55	2.73	2.64	2.22	2.41	2.32	
T ₂ : GA ₃ 50 mg l ⁻¹	0.92	0.91	0.92	2.15	2.24	2.20	1.84	2.03	1.93	
T ₃ : GA ₃ 75 mg l ⁻¹	0.85	0.86	0.86	2.21	2.45	2.33	1.99	2.15	2.07	
T4: NAA 25 mg l ⁻¹	0.85	0.85	0.85	1.56	1.57	1.57	1.31	1.43	1.37	
T ₅ : NAA 50 mg l ⁻¹	0.89	0.94	0.92	2.00	2.30	2.15	1.71	1.73	1.72	
T ₆ : NAA 75 mg l ⁻¹	0.87	0.89	0.88	2.22	2.40	2.31	1.91	2.12	2.02	
T ₇ : GA ₃ 25 mg l ⁻¹ + NAA 25 mg l ⁻¹	0.91	0.86	0.89	2.28	2.23	2.25	1.95	2.16	2.06	
T ₈ : GA ₃ 25 mg l ⁻¹ + NAA 50 mg l ⁻¹	0.94	0.95	0.94	2.19	2.32	2.25	1.90	2.18	2.04	
T ₉ : GA ₃ 25 mg l ⁻¹ + NAA 75 mg l ⁻¹	0.90	0.92	0.91	2.09	2.17	2.13	1.73	1.86	1.80	
T ₁₀ : GA ₃ 50 mg l^{-1} + NAA 50 mg l^{-1}	0.93	0.96	0.95	2.29	2.37	2.33	1.88	2.04	1.96	
T ₁₁ : GA ₃ 75 mg l ⁻¹ + NAA 75 mg l ⁻¹	0.86	0.82	0.84	1.73	1.87	1.80	1.43	1.49	1.46	
T ₁₂ : Control	0.84	0.80	0.82	1.59	1.76	1.68	1.35	1.46	1.41	
Year Mean	0.90	0.90	0.90	2.07	2.20	2.14	1.77	1.92	1.85	
S. Em. ±	0.02	0.03	0.01	0.05	0.06	0.04	0.05	0.05	0.03	
C.D. at 5%	0.06	0.08	0.05	0.16	0.18	0.12	0.13	0.14	0.10	
C.V.%	3.88	4.94	4.44	4.49	4.86	4.69	4.46	4.32	4.39	
YT: S. Em. ±			0.02			0.06			0.05	
YT: C. D. at 5%			NS			NS			NS	

Table 1: Effect of different treatments on chlorophyll content of leaves (mg 100g⁻¹) of onion

Table 2:	Effect of	different	treatments	on reducin	ng sugar,	total	sugar a	and no	n-reducing	g sugar	of onior	l

Turostanonta		Reducing sugar (%)			al sug	ar (%)	Non-reducing sugar (%)			
1 reatments	2018	2019	Pooled	2018	2019	Pooled	2018	2019	Pooled	
T ₁ : GA ₃ 25 mg l ⁻¹	4.67	4.62	4.65	5.69	5.73	5.71	1.02	1.11	1.07	
T ₂ : GA ₃ 50 mg l ⁻¹	4.26	4.63	4.44	5.60	5.95	5.78	1.34	1.32	1.33	
T ₃ : GA ₃ 75 mg l ⁻¹	4.84	6.62	5.73	5.65	7.35	6.50	0.81	0.73	0.77	
T ₄ : NAA 25 mg l ⁻¹	5.21	4.15	4.68	5.80	7.02	6.41	0.59	3.00	1.80	
T ₅ : NAA 50 mg l ⁻¹	4.98	5.24	5.11	6.26	7.55	6.90	1.28	2.31	1.80	
T ₆ : NAA 75 mg l ⁻¹	4.85	3.89	4.37	5.55	8.65	7.10	0.70	4.76	2.73	
T ₇ : GA ₃ 25 mg l ⁻¹ + NAA 25 mg l ⁻¹	4.03	5.15	4.59	5.15	6.52	5.84	1.12	1.37	1.25	
T ₈ : GA ₃ 25 mg l ⁻¹ + NAA 50 mg l ⁻¹	4.09	5.09	4.59	6.60	6.93	6.77	2.51	1.84	2.18	
T ₉ : GA ₃ 25 mg l ⁻¹ + NAA 75 mg l ⁻¹	4.87	4.05	4.46	5.94	8.46	7.20	1.07	4.41	2.74	
T ₁₀ : GA ₃ 50 mg l ⁻¹ + NAA 50 mg l ⁻¹	4.11	5.33	4.72	6.94	6.97	6.95	2.82	1.67	2.25	
T ₁₁ : GA ₃ 75 mg l ⁻¹ + NAA 75 mg l ⁻¹	4.08	4.59	4.34	5.72	5.91	5.82	1.64	1.35	1.49	
T ₁₂ : Control	3.67	4.88	4.28	5.06	7.07	6.07	1.39	2.20	1.79	
Year Mean	4.47	4.85	4.66	5.83	7.01	6.42	1.36	2.17	1.76	
S. Em. ±	0.13	0.13	0.47	0.14	0.20	0.51	0.03	0.06	0.81	
C.D. at 5%	0.37	0.39	NS	0.40	0.59	NS	0.09	0.17	NS	
C.V.%	4.86	4.70	4.78	4.06	4.96	0.17	3.90	4.72	4.62	
YT: S. Em. ±			0.13			0.49			0.05	
YT: C. D. at 5%			0.37			4.63			0.14	

Table 3: Effect of different treatments on	TSS (%) and moisture content (%) of onion
--	---

Tucotmonto		TSS (%)	Moi	ent (%)	
Treatments	2018	2019	Pooled	2018	2019	Pooled
$T_1: GA_3 25 \text{ mg } l^{-1}$	15.20	16.17	15.68	82.10	81.09	81.60
T ₂ : GA ₃ 50 mg l ⁻¹	14.17	12.67	13.42	82.73	79.15	80.94
T ₃ : GA ₃ 75 mg l ⁻¹	13.43	14.00	13.72	83.49	79.50	81.50
T4: NAA 25 mg l ⁻¹	14.70	15.00	14.85	79.26	78.90	79.08
T ₅ : NAA 50 mg l ⁻¹	14.37	15.00	14.68	84.36	82.62	83.49
T ₆ : NAA 75 mg l ⁻¹	14.40	15.58	14.99	84.89	83.07	83.98
T ₇ : GA ₃ 25 mg l ⁻¹ + NAA 25 mg l ⁻¹	14.87	15.33	15.10	78.81	81.03	79.92
T ₈ : GA ₃ 25 mg l ⁻¹ + NAA 50 mg l ⁻¹	13.87	15.75	14.81	82.68	82.71	82.69
T ₉ : GA ₃ 25 mg l^{-1} + NAA 75 mg l^{-1}	15.03	15.33	15.18	80.86	80.56	80.71
T ₁₀ : GA ₃ 50 mg l^{-1} + NAA 50 mg l^{-1}	13.87	15.30	14.58	82.03	81.14	81.59
T ₁₁ : GA ₃ 75 mg l^{-1} + NAA 75 mg l^{-1}	14.73	14.83	14.78	81.77	82.97	82.37
T ₁₂ : Control	13.73	15.00	14.37	79.16	78.47	78.81
Year Mean	14.36	15.00	14.68	81.85	80.93	81.39
S. Em. ±	0.31	0.38	0.43	1.33	1.09	0.86
C.D. at 5%	0.92	1.12	NS	3.89	3.18	2.44
C.V.%	3.80	4.42	4.14	2.81	2.32	2.58
YT: S. Em. ±	0.35			1.21		
YT: C. D. at 5%			1.00			NS

Table 4: Effect of different treatments on ascorbic acid (mg 100g⁻¹) and bulb pH of onion

Treatmente	Asco	rbic acid (m	Bulb pH			
Treatments	2018	2019	Pooled	2018	2019	Pooled
T ₁ : GA ₃ 25 mg l ⁻¹	10.41	10.50	10.45	4.48	4.61	4.55
T ₂ : GA ₃ 50 mg l ⁻¹	8.40	8.41	8.40	4.63	4.60	4.62
T ₃ : GA ₃ 75 mg l ⁻¹	8.47	8.52	8.49	4.77	4.72	4.75
T4: NAA 25 mg l ⁻¹	8.67	8.89	8.78	4.60	4.59	4.59
T ₅ : NAA 50 mg l ⁻¹	9.24	9.27	9.25	4.67	4.74	4.71
T ₆ : NAA 75 mg l ⁻¹	8.79	8.52	8.66	4.74	4.77	4.76
T ₇ : GA ₃ 25 mg l ⁻¹ + NAA 25 mg l ⁻¹	8.28	9.61	8.95	4.65	4.65	4.65
T ₈ : GA ₃ 25 mg l ⁻¹ + NAA 50 mg l ⁻¹	9.43	9.45	9.44	4.67	4.52	4.59
T ₉ : GA ₃ 25 mg l^{-1} + NAA 75 mg l^{-1}	8.74	8.63	8.68	4.68	4.59	4.64
T_{10} : GA ₃ 50 mg l ⁻¹ + NAA 50 mg l ⁻¹	9.81	9.38	9.60	4.62	4.67	4.65
T_{11} : GA ₃ 75 mg l ⁻¹ + NAA 75 mg l ⁻¹	8.46	8.40	8.43	4.78	4.70	4.74
T ₁₂ : Control	8.10	8.22	8.16	4.76	4.38	4.57
Year Mean	8.90	8.98	8.94	4.67	4.63	4.65
S. Em. ±	0.21	0.26	0.18	0.06	0.07	0.07
C.D. at 5%	0.62	0.75	0.50	0.17	0.20	NS
C.V.%	4.10	4.93	4.54	2.12	2.58	2.36
YT: S. Em. ±			0.23			0.06
YT: C. D. at 5%			NS			0.18

References

- 1. Arteca RN. Plant growth substances: Principles and applications. CBS Publishers, New Delhi, 1996.
- Chaudhary BR, Sharma MD, Shakya SM, Gautam DM. Effect of plant growth regulators on growth, yield and quality of chilli (*Capsicum annuum* L.) at Rampur, Chitwan. J. Inst. Agric. Anim. Sci. 2006; 27:65-68.
- 3. Gomatinayagam M, Anuradha VE, Zhao C, Ayoola GA, Jaleel CA, Annerselvam RP. ABA and GA₃ affect the growth and pigment composition in *Andrographis paniculata* Wall. Frontiers Biol. China. 2000; 4:337-341.
- Govind, Maji S, Kumawat R, Pal A, Kumar S, Saha S. Improvement of growth, yield and quality of garlic (*Allium sativum* L.) cv. G-282 through a novel approach. Bioscan. 2015; 10(1):23-27.
- Ouzounidou G, Giannakoula A, Asfi M, Ilias I. Differential responses of onion and garlic against plant growth regulators. Pakistan J. Bot. 2011; 43(4):2051-2057.
- 6. Ouzounidu G, Ilias I. Hormone induced protection of sunflower photosynthetic apparatus against Cu toxicity. Biologia Plantarum. 2005; 49:223-228.
- 7. Ouzounidu G, Ilias I, Giannakoula A, Parthena P. Comparative study on the effects of various plant growth regulators on growth, quality and physiology of (*Capsicum annum* L.). Pakisthan J. Bot. 2010; 42(2):805-814.
- 8. Patel MJ, Patel HC, Chavda JC. Influence of plant growth regulators and their application methods on yield and quality of onion (*Allium cepa* L.). Asian J. Hort. 2010; 5(2):263-265.
- 9. Purohit SS. Growth Regulators, Plant Physiology, National Book Trust, New Delhi, 2007, 32-46.
- 10. Shah SH, Ahmad I, Saimullah. Respones of *Nigella sativa* to foliar application of gibberellic acid and kinetin. Biologia Plantarum, 2007; 51:563-566.
- Tiwari RS, Ankur A, Sengar SC. Effect of bioregulators on growth, bulb yield, quality and storability of onion cv. Pusa Red. Indian J. Pl. Physiol. 2003; 8(4):411-413.
- Weaver RJ. Plant Growth Substances in Agriculture. W. H. Freeman and Co., San Francisco, 1972, 339p.