

International Journal of Chemical Studies

P-ISSN: 2349–8528 E-ISSN: 2321–4902 www.chemijournal.com IJCS 2020; 8(6): 3030-3034 © 2020 IJCS Received: 17-09-2020 Accepted: 23-10-2020

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Department of Plant Pathology, Vasantrao Naik Marathwada Agriculture University, Parbhani, Maharashtra, India Induced systemic resistance against X. axonopodis pv. citri. through chemical elicitors

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DOI: https://doi.org/10.22271/chemi.2020.v8.i6aq.11982

Abstract

ISR elicitors *viz.*, Salicylic acid, B-aminobutyric acid, Jasmonic acid, Isonicotinic acid, L-arginine, Lmethionine and L-ornithine were evaluated *in vitro* (Pot culture), observations were recorded on Per cent disease intensity and lesion diameter of acid lime bacterial canker. The ISR elicitors (chemical) applied plants of acid lime revealed that, the mean per cent disease intensity, in all the treatments it was decreased over control. However, minimum mean per cent disease intensity was recorded with Baminobutyric acid (08.00) followed by Salicylic acid (08.66), L-methionine (09.89), Jasmonic acid (13.11), Isonicotinic acid (17.78) and L-ornithine (19.66); whereas it was recorded maximum in Larginine (19.88). After 15 days of challenge inoculation with *X. axonopodis* pv. *citri*, the lesion diameter decreased over control. However, no lesson was recorded with Salicylic acid and B-aminobutyric acid which were found significantly superior amongst the rest all; whereas, minimum lesion diameter was recorded with L-methionine (01.33). Decreasing trend in Lesion diameter after 30 days and 45 days of challenge inoculation with *X. axonopodis* pv. *citri*, over control was observed. However, minimum lesion diameter after 30 days and 45 days was recorded same with Salicylic and B-aminobutyric acid (01.33 and 02.67, respectively) and found superior over the rest all; whereas, it was found maximum with L-arginine (05.33 and 07.00, respectively) and L-ornithine (05.33 and 07.67, respectively).

Keywords: Bacterial canker, acid lime, chemicals, induced systemic resistance

Introduction

Copper (Cu) based bactericides are a standard control measure for citrus canker worldwide (Kuhara, 1978., Stall et al. 1982., Leite and Mohan 1990) [27, 29, 28]. Cu reduces bacterial populations on leaf surfaces, but multiple applications are needed to achieve adequate control on susceptible citrus hosts (Graham, 2001., Stall et al., 1982 & 1990)^[30, 29]. Long-term use of Cu bactericides have other possible disadvantages, including resistance in xanthomonad populations (Marco, and Stall 1983., Rinaldi and Leite 2000) [32, 30, 33]. Systemic acquired resistance (SAR) is a mechanism of induced defense that may confer long-lasting protection against a broad spectrum of microorganisms (van Loon et al. 2006)^[22]. Induced resistance requires the signal molecule salicylic acid and is associated with the accumulation of pathogenicity-related (PR) proteins, and induction of defense enzymes. The latter two are thought to contribute to resistance. (Ojha and Chaterjee, 2012)^[23]. β-Aminobutyric acid (BABA) is a non-protein amino acid which induces resistance against a broad range of disease-causing organisms including fungi, bacteria, viruses, and nematodes (Jakab et al. 2001; Francis et al. 2009) ^[24, 25]. Pretreatment with BABA can also be effective against bacterial diseases by inducing systemic resistance against X. citri pv. citri (Graham and Leite 2007; Francis et al. 2009) [25]. Resistance to diseases in plants was induced by BABA, either through the activation of a signaling pathway that depends on SA or through the activation of a novel signaling cascade not dependent on SA but on jasmonic acid or ethylene (Zimmerli et al. 2000) [26].

Induced resistance can also be provoked by some chemicals such as salicylic acid (SA), jasmonic acid (JA), Iso-nicotinic acid (NA) (Esmailzadeh *et al.* 2008., Wang and Liu 2012., Li and Wang 2013) ^[12, 13, 14]. Salicylic acid plays an important role in induction of plant defense against a variety of biotic and abiotic stresses through morphological, physiological and biochemical mechanisms (Yao and Tian, 2005., Hayat *et al.* 2010) ^[15, 16]. It is involved in endogenous signaling, mediating in plant defense against pathogens. Jasmonic acid is a class of lipidic plant hormones, not only involved in plant-microbe's interactions in defense and symbiosis (Ballaré, 2011) ^[17] but also induced secondary plant metabolites, like alkaloids,

Corresponding Author: Jadhav RR Department of Plant Pathology, Vasantrao Naik Marathwada Agriculture University, Parbhani, Maharashtra, India terpenoids, flavonoids, coumarins and so forth (Faurie *et al.* 2009) ^[18]. In addition, NA is involved in metabolic functions such as biosynthesis of nucleic acids, amino acids and pantothenate, lignin formation and synthesis of pathogenesis-related (PR) proteins and regulated redox process (Hanson and Roje 2001) ^[19]. So far, some studies have provided the evidence that phytohormones like SA, JA and NA have the potentiality for mitigating plant disease by modulating redox balance activating enzymes (Cossins., 2000., Wen *et al.* 2008) ^[20, 21]. Above information reveals that chemicals like salicylic acid, β -aminobutyric acid (BABA), jasmonic acid, Isonicotinic acid and L-arginine, L-methionine, L-ornithine create a favorable condition for plant to resist pathogen by modulating physiological and molecular process.

However, the research on the effect of ISR chemicals on the induction of resistance in plants is very limited. So, we tested the effect of the salicylic acid (SA), β -aminobutyric acid (BABA), jasmonic acid, Isonicotinic acid and L-arginine, L-methionine, L-ornithine on induced resistance against bacterial canker disease in acid lime plants. The term ISR used here to include ISR and SAR inducers with activity against pathogen.

Material and methods ISR Chemicals

ISR Elicitors chemicals *viz.*, salicylic acid (SA), β aminobutyric acid (BABA), jasmonic acid, Isonicotinic acid and L-arginine, L-methionine, L-ornithine were used during present study. One year acid lime (*Citrus aurantifoliia*) healthy seedlings were collected from Central nursery VNMKV, Parbhani and was transplanted in polyethylene bags/ earthen pots (one seedling/bag or pot) containing sterilized potting mixture of sterile loamy soil and sand (2:1). Potted acid lime seedlings were maintained under shed/ in screen house and watered as and when required. Acid lime plants were foliar spry-treated with ISR @ 50 ppm concentration of each. Distilled water spry-treated plants were also maintained as control. This experiment was designed in screen house (Pot culture) by using CRD with three replications and total eight treatments including control.

Bacterial culture and inoculation

For the preparation of the bacterial suspension, the bacterial strain was cultured in Nutrient agar (NA) and grown at 28°C for 24 h. Then, a single colony was transferred to Yeast Extract Peptone medium and grown at 28°C for 24 h to log phase. The final bacterial suspension was pelleleted at 10,000 rpm for 20 min and again suspended in distilled water to reach to a 7×10^8 colony forming unit (CFU/ml).

Immature leaves were inoculated using a tuberculin syringe to produce a zone of water-soaked tissue. The infiltrated areas of the leaf were approximately 6 mm diameter and contain approximately 5 μ l of bacterial suspension. Two injection infiltrations were performed on each side of the mid-vein and eight leaves were inoculated per plant.

Observations of per cent disease intensity and lesion diameter were recorded at 15, 30 and 45 days after challenge inoculation of bacterial suspension. The results of experiments are mentioned in table

Plant material and treatments

Table 1: In vitro efficacy	v of ISR elicitors	(Chemicals) on	ner cent disease	intensity (pot culture)
LADIC L. III VIIIO CIIICac	y of ion chemory	Chemicals) on	per cent discuse	mensity (por culture)

Tr. No.	Tracetore		PDI	Mean PDI	Maar BDC		
	Treatments	15DAI	30DAI	45DAI	Mean PDI	Mean PDC	
T1	Jasmonic acid	3.00 (9.87)*	13.67 (21.67)*	22.67 (28.41)*	13.11	47.09	
T ₂	Salicylic acid	0.00 (0.00)	8.33 (16.76)	17.67 (24.83)	08.66	65.05	
T ₃	Isonicotinic acid	9.00 (17.43)	17.67 (24.83)	26.67 (31.07)	17.78	28.24	
T_4	B-aminobutyric acid (BABA)	0.00 (0.00)	8.67 (17.10)	15.33 (23.27)	08.00	67.71	
T ₅	L-arginine	9.33 (17.77)	21.00 (27.24)	29.33 (32.77)	19.88	19.77	
T ₆	L-methionine	2.67 (9.26)	8.33 (16.76)	18.67 (25.57)	09.89	60.08	
T7	L-ornithine	9.00 (17.43)	21.00 (27.23)	29.00 (32.56)	19.66	20.66	
T8	Control	12.67 (20.82)	25.67 (30.41)	36.00 (36.85)	24.78	00.00	
	S.E. <u>+</u>	0.64	0.66	0.58			
	C.D. (P=0.01)	1.95	2.00	1.76			
*Mean of three replications Fingers in parenthesis are angular transformed values							

Results (Table 1 and Fig. 1) indicated that in ISR elicitor applied plants of acid lime after 15 days of challenge inoculation with X. axonopodis pv. citri, percent disease intensity was significantly decreased over control and it was ranged from 0.00 to 09.33. However, disease intensity was recorded with Salicylic acid and B-aminobutyric acid which were found superior amongst the rest all; whereas minimum per cent disease intensity was recorded in L-methionine (02.67) followed by Jasmonic acid (03.00), both were at par with each other, while L-arginine recorded maximum per cent disease intensity which was at par with remaining two treatments viz., Isonicotinic acid (09.00) and L-ornithine (09.00). After 30 days of challenge inoculation with X. axonopodis pv. citri, in all the treatments percent disease intensity significantly decreased over control and it was ranged from 08.33 to 25.67. However, minimum per cent disease intensity was recorded with Salicylic acid (08.33) followed by L-methionine, B-aminobutyric acid (08.67), but all three were at par with each other and found significantly superior treatments, Isonicotinic acid (17.67), Jasmonic acid (13.67); whereas least superior treatments were L-arginine (21.00) and L-ornithine (21.00) which recoded maximum per cent disease intensity but both were at par with each other. The percent disease intensity after 45 days of challenge inoculation with X. axonopodis pv. citri, also significantly decreased over control and it was ranged from 15.33 to 29.33. However, significantly superior treatment was found Baminobutyric acid (15.33) in which minimum per cent disease intensity was recorded, followed by Salicylic acid (17.67), Lmethionine (18.67) but both were at par with each other, Jasmonic acid (22.67), Isonicotinic acid (26.67), L-ornithine (29.00) and L-arginine (29.33), but Isonicotinic acid, Lornithine and L-arginine were at par with each other and found least superior.

In case of mean per cent disease intensity, in all the treatments it was decreased over control (24.78) and varied from 08.00 to

19.88. However, minimum mean per cent disease intensity was recorded with B-aminobutyric acid (08.00), followed by Salicylic acid (08.66), L-methionine (09.89), Jasmonic acid (13.11), Isonicotinic acid (17.78) and L-ornithine (19.66) while maximum mean per cent disease intensity was recorded in L-arginine (19.88). Mean per cent disease control was

ranged from 19.77 to 67.71. However, maximum mean per cent disease control was recorded with B-aminobutyric acid (67.71), followed by Salicylic acid (65.05), L-methionine (60.08), Jasmonic acid (47.09), Isonicotinic acid (28.24) and L-ornithine (20.66). While, minimum mean per cent disease control was recorded with L-arginine (19.77).

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Tr.	Treatments	Lesion diameter (mm)*			Mean lesion	Per cent reduction of lesion	
No.	1 reatments	15 DAI	30 DAI	45 DAI	diameter (mm)	diameter over control	
T1	Jasmonic acid	1.67 (1.62)*	2.67 (1.91)*	4.33 (2.30)*	2.89	52.70	
T_2	Salicylic acid	0.00 (1.00)	1.33 (1.52)	2.67 (1.91)	1.33	78.23	
T ₃	Isonicotinic acid	1.67 (1.62)	3.00 (2.00)	4.67 (2.37)	2.55	58.26	
T 4	B-aminobutyric acid (BABA)	0.00 (1.00)	1.33 (1.52)	2.67 (1.91)	1.66	72.83	
T5	L-arginine	2.67 (1.91)	5.33 (2.50)	7.00 (2.82)	5.00	18.00	
T ₆	L-methionine	1.33 (1.52)	2.00 (1.71)	3.00 (1.98)	2.11	65.45	
T 7	L-ornithine	2.33 (1.82)	5.33 (2.51)	7.67 (2.94)	5.11	16.36	
T8	Control	3.67 (2.15)	6.00 (2.64)	8.67 (3.10)	6.11	00.00	
	S.E. <u>+</u>	0.08	0.10	0.08			
	C.D. (P=0.01)	0.25	0.30	0.27			
*Mean of three replications							
Fingers in parenthesis are square root transformed values							

Results (Table 2, Fig. 2) revealed that, after 15 days of challenge inoculation with X. axonopodis pv. citri, the lesion diameter significantly decreased over control (03.67) and it was ranged from 0.00 to 02.67. However, no lesson was recorded with Salicylic acid and B-aminobutyric acid which were found significantly superior amongst the rest all; whereas, minimum lesion diameter was recorded with Lmethionine (01.33), followed by Jasmonic acid (01.67), Isonicotinic acid (01.67) but were at par with each other, while with L-ornithine (02.33) and L-arginine (02.67) recorded maximum lesion diameter which were at par with each other. Lesion diameter after 30 days of challenge inoculation with X. axonopodis pv. citri, also significantly decreased over control (06.00) and it was ranged from 1.33 to 5.33. However, minimum lesion diameter was recorded with Salicylic (01.33) acid and B-aminobutyric acid (0.133), but were at par with each other and found significantly superior over the rest all followed by L-methionine (02.0), Jasmonic acid (01.67), Isonicotinic acid (03.00), but were at par with each other; whereas, maximum lesion diameter was found with L-arginine (05.33) and L-ornithine (05.33), but were at par. Lesion diameter after 45 days of challenge inoculation with X. axonopodis pv. citri, also significantly decreased over control (08.67) and it was ranged from 02.67 to 07.67. However, minimum lesion diameter was recorded with Salicylic (02.67) acid and B-aminobutyric acid (02.67), but were at par with each other and found significantly superior over the rest all; followed by L-methionine (03.00), Jasmonic acid (04.33), Isonicotinic acid (04.67), but were at par with each other; whereas, maximum lesion diameter was found with L-arginine (07.00) and L-ornithine (07.67).

Mean lesion diameter, in all the treatments was decreased over control (06.11) and varied from 01.33 to 05.11. However, minimum mean lesion diameter was recorded with Salicylic acid (01.33), followed by B-aminobutyric acid (01.66), L-methionine (02.11), Isonicotinic acid (02.55), Jasmonic acid (02.89), and L-arginine (05.00) while, maximum mean lesion diameter was recorded with Lornithine (05.11); whereas mean per cent reduction of lesion diameter over control (0.00) was ranged from 16.36 to78.23. However, maximum mean per cent reduction of lesion diameter over control was recorded with Salicylic acid (78.23), followed by B-aminobutyric acid (72.83), Lmethionine (65.45), Isonicotinic acid (58.26), Jasmonic acid (52.70), and L-arginine (18.00) while, minimum mean per cent reduction of lesion diameter over control was recorded in L-ornithine (13.36).

Similar findings were recorded earlier by many workers. Beheshti et al. (2011) treated ISR chemicals, by spraytreatment on the lime (Citrus aurantifolia) plants challenge inoculated with X. axonopodis pv citri. with β -Aminobutyric Acid (BABA) @ 250 ppm, salicylic acid @ 2 mM. Lesion diameters of inoculated leaves were evaluated twenty days after treatment inoculated citrus leaves with Xcc. Results revealed that, spray treatment with BABA reduced lesion size by about 50% as compared to the water control. Thus while water treated leaves showed lesion sizes of about 1.2 cm, BABA and treated plants had lesions of only about 0.6 cm. While, significant reduction in lesion size was observed following treatment with salicylic acid. Hasabi et al. (2014)^[9] studied effect of three amino acids viz., L-arginine, Lmethionine, L-ornithine, and distilled water application on induced resistance against citrus canker caused by X. axonopodis pv. citri. in lime plants. Plants were inoculated with the suspension of the test bacterium and canker lesion diameter was measured four weeks after inoculation. They reported that plants treated L-methionine expressed significantly increased induced resistance and decreased disease severity, by reducing necrotic lesion size by about 78.5% as compared to untreated control. No significant reduction in lesion size was observed following treatment with L-arginine or L-ornithine.

Conclusion

All the ISR elicitors (chemicals) significantly reduced disease intensity as well as lesion diameter on acid lime. Of all the chemicals evaluated, B-aminobutyric acid (BABA) was found most superior; second best was Salicylic acid.

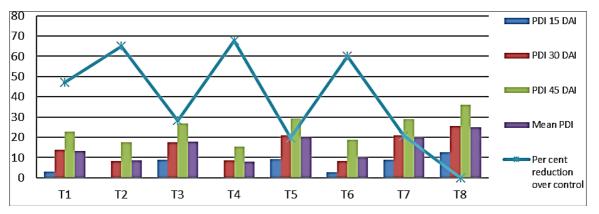


Fig 1: In vitro efficacy of ISR elicitors (Chemicals) on per cent disease intensity (pot culture)

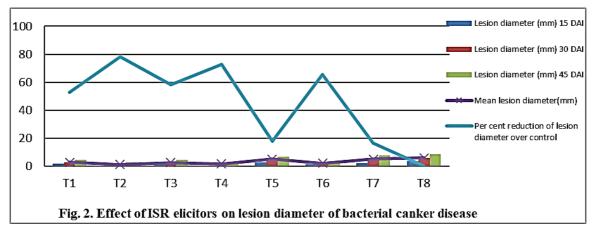


Fig 2: Effect of ISR elicitors on lesion diameter of bacterial canker disease

References

- 1. Anonymous. Indian Horticulture Data Base. National Horticulture Board, 2018, 42-67.
- Beheshti B, Sharifi-Sirchi GR, Mansouri M, Hosseinipour A, Schlaich NL. Resistance to citrus canker in key/Mexican lime induced by β-aminobutyric acid and green tea. American J. Agril. Biol. Sci 2011;6(2):242-248.
- Civerolo EL. Bacterial canker disease of citrus. J. Rio Grande Valley Hort. Soc 1984;37:127-146.
- 4. Das AK. Citrus canker A review. J. Appl. Hort 2003;5(1):52-60.
- Das R, Mondal B, Mondal P, Khatua DC, Mukherjee N. Biological management of citrus canker on acid lime through *Bacillus subtilis* (S-12) in West Bengal. Indian. J. Biopest 2014;7:38-41.
- 6. Fawcett HS, Jenings AE. Records of citrus canker from herbacium specimens of the genus Citrus in England and the United States. Phytopath 1993;23:820-824.
- 7. Gottwald TR, Graham JH, Schubert TS. Citrus canker the pathogen and its impact. Plant Helth Progress, 2002, 812-01RV.
- Gottwald TR, Graham JH, Civerolo EL, Barret HC, Hearn CJ. Differential host range reaction of citrus and citrus relatives to citrus canker and citrus bacterial spot determined by leaf measophyl susceptibility. Pl. Dis 1993;77:1004-1009.
- 9. Hasabi V, Askari H, Mehdi AS, Zamanizadeh H. Effect of amino acid application on induced resistance against citrus canker disease in lime plants. J. Pl. Prot. Res 2014;54:144-149.

- 10. Stall RE, Civerolo EL. Research relating to the recent outbreak of citrus canker in Florida. Ann. Rev. Phytopath 1991;29:399-420.
- Verniere C, Hartung JS, Pruvost OP, Civerolo EL, Alvsrez AM, Maestri P *et al.* Characterization of phenotypically distinct strains of *Xanthomonas axonopodis* pv.*citri.* from South west Asia. European. J. Pl. Path 1998;104(5):477-487.
- 12. Esmailzadeh M, Soleimani MJ, Rouhani H. Exogenous applications of salicylic acid for inducing systemic acquired resistance against tomato stem canker disease. J. Biological Sci 2008;8:1039-1044.
- 13. Wang Y, Liu JH. Exogenous treatment with salicylic acid attenuatesc occurrence of citrus canker in susceptible navel orange (*Citrus sinensis* Osbeck). J. Pl. Physio 2012;169:1143-1144.
- 14. Li J, Wang N. Foliar application of biofilm formationinhibiting compounds enhances control of citrus canker caused by *Xanthomonas citri* subsp. *citri*. Phytopathol 2013;104:134-142.
- 15. Yao H, Tian S. Effects of pre- and post-harvest application of salicylic acid or methyl jasmonate on inducing disease resistance of sweet cherry fruit in storage. Postharvest Biol. and Tech 2005;35:253-262.
- 16. Hayat Q, Hayat S, Irfan M, Ahmad A. Effect of exogenous salicylic acid under changing environment: a review. Environ. and Experimental Bot 2010;68:14-25.
- Ballaré CL. Jasmonate-induced defenses: a tale of intelligence, collaborators and rascals. Trends Pl. Sci 2011;16:249-257.
- 18. Faurie S, Cluzet MF, Marie-France CC, Merillon JM. Methyl jasmonate/ ethephon co-treatment synergistically induces stilbene production in vitis vinifera cell

suspensions but fails to trigger resistance to erysiphe necator. J, Int. des Sci 2009;43:99-110.

- 19. Hanson AD, Roje S. One-carbon metabolism in higher plants. Ann.Rev. Pl. Physio. and Pl. Mol. Biol 2001;52:119-137.
- 20. Cossins EA. The fascinating world of folate and one-carbon metabolism. Canadian J. Bot 2000;78:691-708.
- 21. Wen PF, Chen JY, Wan SB, Kong WF, Zhang P *et al.* Salicylic acid activates phenylalanine ammonia-lyase in grape berry in response to high temperature stress. Pl. Growth Regulation 2008;55:1-10.
- 22. Van Loon LC, Rep M, Pieterse CMJ. Significance of inducible defense-related proteins in infected plants. Annu. Rev. Phytopathol 2006;44(1):135-162.
- 23. Ojha S, Chandra Chatterjee N. Induction of resistance in tomato plants against *F. oxysporum* f. sp. *Lycopersici* mediated through salicylic acid and *T. harzianum*. J. Plant Prot. Res 2012;52(2):220-225.
- Jakab G, Cottier V, Toquin V, Rigoli G, Zimmerli L. β-Aminobutyric acidinduced resistance in plants. Eur. J. Plant Pathol 2001;107(1):29-37.
- 25. Francis MI, Redondo A, Burns JK, Graham JH. Soil application of imidacloprid and related SAR-inducing compounds produces effective and persistent control of citrus canker. Eur. J. Plant Pathol 2009;124(2):283-292.
- 26. Zimmerli L, Jakab G, Matraux JP, Mauch-Mani B. Potentiation of pathogen-specific defense mechanisms in *Arabidopsis* by β -aminobutyric acid. Proc. Natl. Acad. Sci. USA 2000;97(23):12920-12925.
- 27. Kuhara S. Present epidemic status and control of the citrus canker disease (*Xanthomonas citri* (Hasse) Dowson) in Japan. Rev. Plant Prot. Res 1978;11:132-142.
- 28. Leite RPJr, Mohan SK. Integrated management of the citrus bacterial canker disease caused by *X. campestris* pv. *citri* in the State of Paraná, Brazil. Crop Prot 1990;9:3-7.
- 29. Stall RE, Miller JW, Marco GM, Canteros de Echenique BI. Timing of sprays to control cancrosis of grapefruit in Argentina. Proc. Int. Soc. Citric 1982;1:414-417.
- 30. Graham JH. Varietal susceptibility to citrus canker: Observations from southern Brazil. Citrus Ind 2001;82(6):15-17.
- Stall RE, Miller JW, Marco GM, Canteros de Echenique BI. Population dynamics of *Xanthomonas citri* causing cancrosis of citrus in Argentina. Proc. Fla. State Hortic. Soc 1980;93:10-14.
- 32. Marco GM, Stall RE. Control of bacterial spot of pepper initiated by strains of *Xanthomonas campestris* pv. *vesicatoria* that differ in sensitivity to copper. Plant Dis 1983;67:779-781.
- 33. Rinaldi DAMF, Leite RPJr. Adaptation of *Xanthomonas axonopodis* pv. *citri* population to the presence of copper compounds in nature. Proc. Int. Soc. Citric 2000;2:1064.